



**5th HALLE  
CONFERENCE**  
ON RECOMBINANT PROTEINS

19th – 20th  
FEB 2015  
LEOPOLDINA



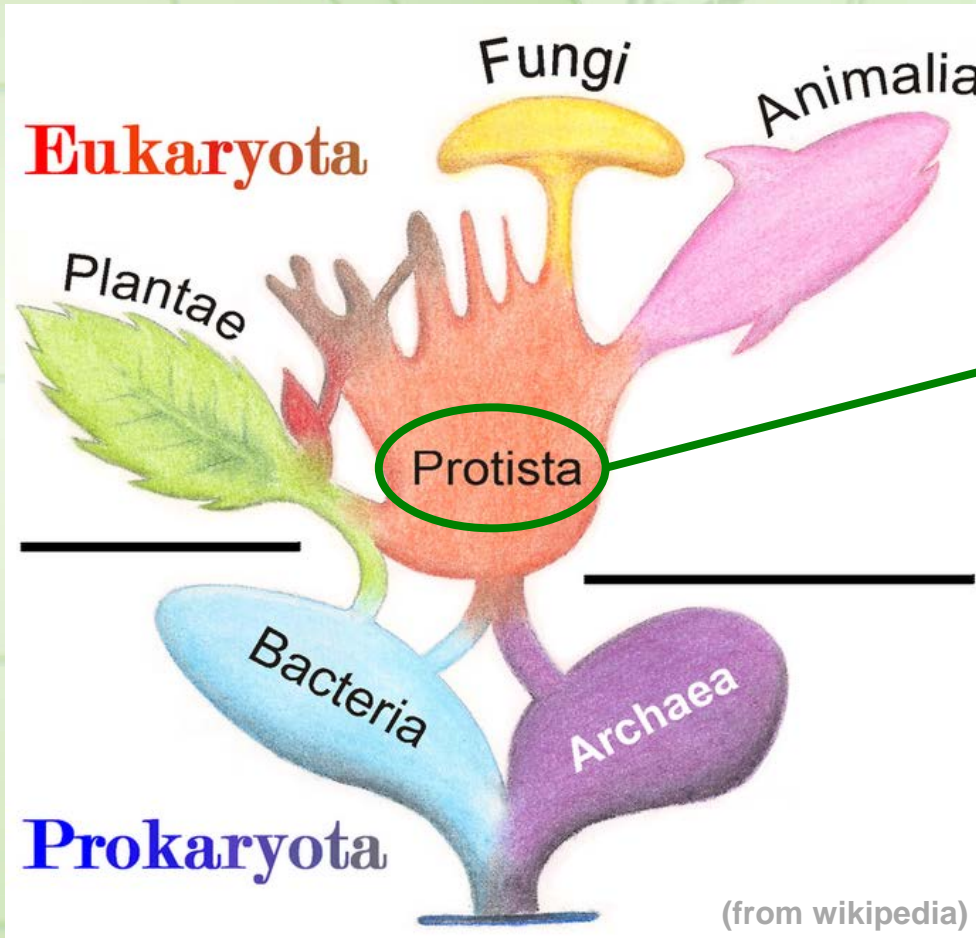
**LEXSY taking off:  
Selected examples for protein production  
with *Leishmania tarentolae***

Jena Bioscience GmbH  
Loebstedter Str. 80  
07749 Jena, Germany  
Tel.: +49-3641-628-5000  
Fax: +49-3641-628-5100

e-Mail: [expression@jenabioscience.com](mailto:expression@jenabioscience.com)  
<http://www.jenabioscience.com/LEXSY>

Time warp to 1998

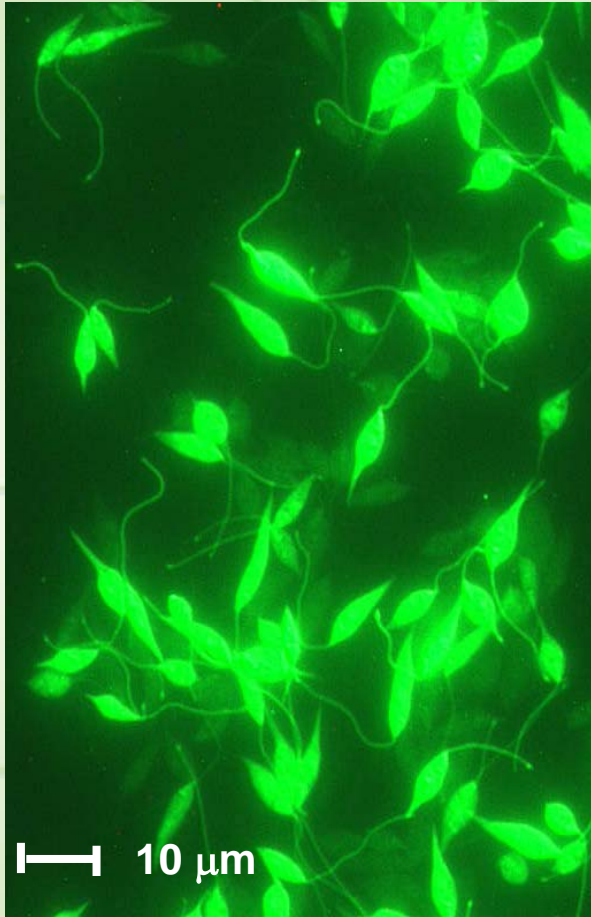
Why not an expression system that **combines** prokaryotic easiness with eukaryotic capabilities?



- Large, diverse kingdom of organisms
- Many didn't look very promising...
- ...but one did:

***Leishmania tarentolae***

# So *Leishmania tarentolae* became LEXSY



## *Leishmania tarentolae*

- Nonpathogenic to mammals (S1-clearance)
- Very happy in culture (flasks and fermenters)
- Complete eukaryotic protein synthesis / folding / modification machinery (PTMs<sup>(1)</sup>)

Not even 10 years of hard  
lab work...

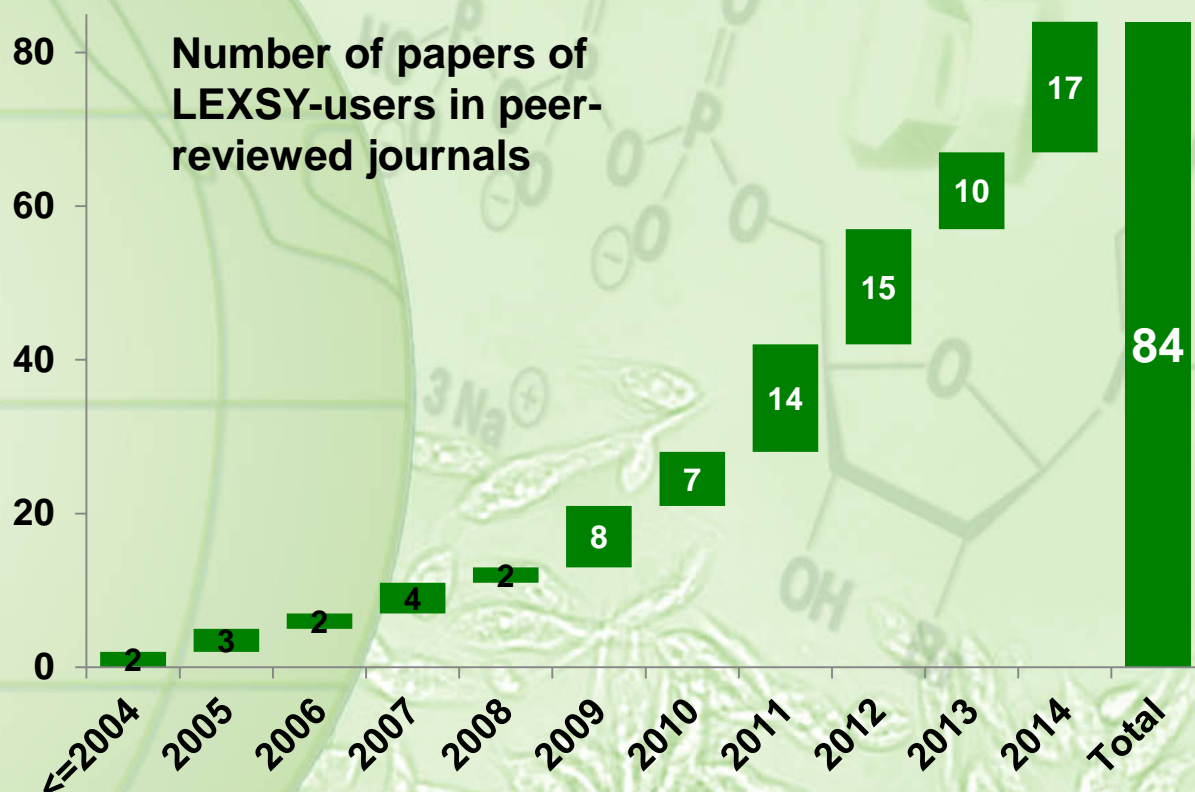
## LEXSY

- Suitable for many *types* of proteins including membrane, cytosolic, nuclear and secreted proteins
- Approx. 80% positive expression projects with yields up to 500 mg per litre of culture
- Cell-free version (*in vitro* translation with LEXSY extracts)

(1) PTMs = post-translational modifications such as mammalian-type glycosylation, phosphorylation, glypiation (GPI anchoring), acetylation, prenylation, myristoylation, ADP-ribosylation, proteolytic processing, oligomerisation...

# Soon more and more people started using LEXSY

To date, LEXSY is established in roughly 200 labs worldwide



Exponential take-off...? Last few minutes some selected LEXSY examples

# LEXSY makes antigens, EPO, interleukin and interferon

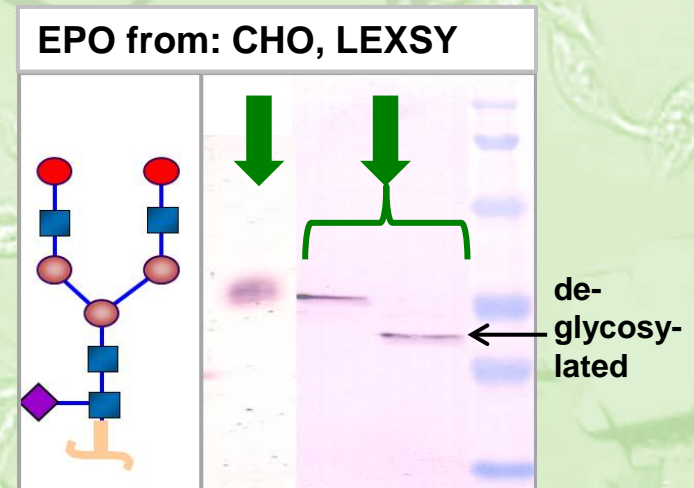
## Vaccines

- Production of recombinant viral antigens (influenza<sup>(1)</sup>, Hepatitis <sup>(2)</sup>, Papilloma Virus<sup>(3)</sup>)
- Production of recombinant parasite antigens (Leishmaniasis<sup>(4)</sup>)
- Whole LEXSY cells as live vaccine delivery vector in tumor models<sup>(5,6)</sup>

## Cytokines

- Human interferon gamma (IFN $\gamma$ ) with yields of 10 mg/L<sup>(8)</sup>
- N-glycosylated, antivirally active human interleukin 29 (IL-29<sup>(9)</sup>)
- Mammalian-type N-glycosylated homogeneous biologically active human EPO<sup>(7)</sup>

- (1) Pion *et al.* (2014) *Vaccine* 32: 5570
- (2) Baechlein *et al.* (2013) *Journal of Virological Methods* 193: 238
- (3) Hosseinzadeh *et al.* (2013) *Drug Delivery* 20: 190
- (4) Chamakh-Ayari *et al.* (2014) *PLOS ONE* 9: e92708
- (5) Saljoughian *et al.* (2013) *PLoS Neglected Tropical Dis.* 7: e2174
- (6) Salehi *et al.* (2012) *Immunotherapy* 4: 1107
- (7) Breitling *et al.* (2002) *Prot Expr and Purification* 25: 209
- (8) Davoudi *et al.* (2011) *World J. Microbiol. Biotechnology* 27: 1893
- (9) Taramchi *et al.* (2013) *Iranian Journal of Biotechnology* 11: 168

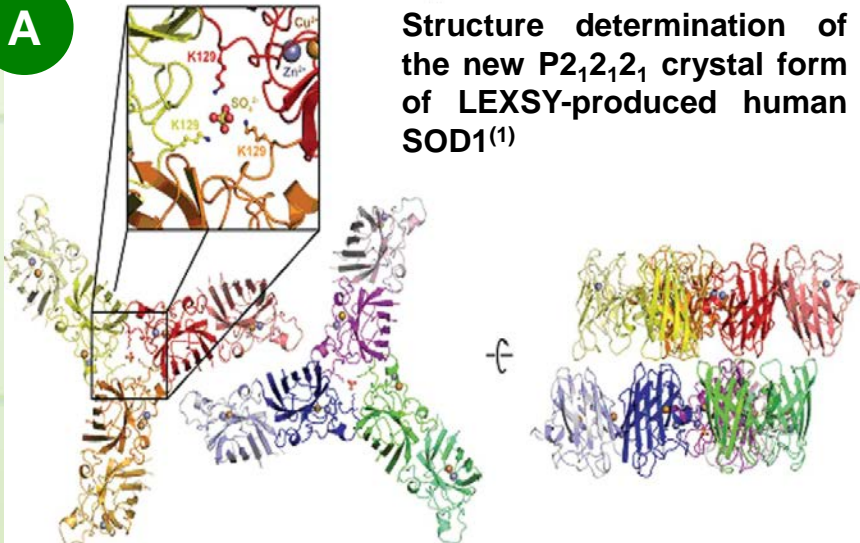


# LEXSY-proteins can be crystallized and NMRed

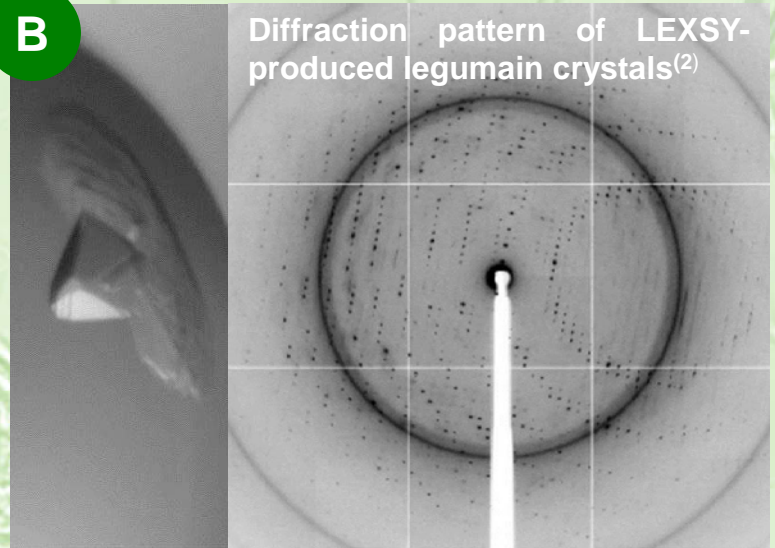
## Structural biology

- Crystal structure of LEXSY-made antioxidant human Cu/Zn superoxide dismutase SOD1<sup>(1)</sup> (A)
- LEXSY-made legumain (cyst. protease) crystals diffract to 2.5 Å<sup>(2)</sup> (B)
- <sup>15</sup>N-HSQC NMR yields assignment of 18x <sup>15</sup>N-Val in a 28 kDa protein<sup>(3)</sup>

A



B

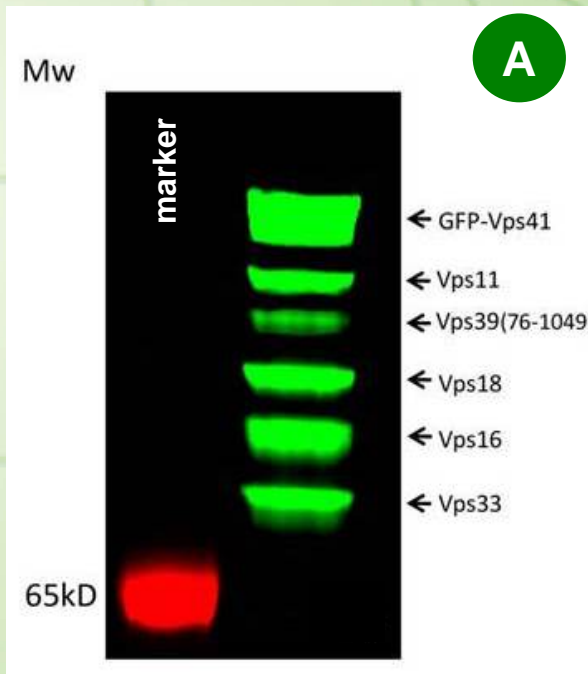


- (1) Gazdag *et al.* (2010) *Acta Crystallographica* F66: 871  
(2) Dall & Brandstetter (2012) *Acta Crystallographica* F68: 24  
(3) Niculae *et al.* (2006) *Protein Expression and Purification* 48: 167

# *In vitro* LEXSY (cell-free expression based on LEXSY cell extracts) is an alternative to traditional wheat germ or RRL systems

## *In vitro* LEXSY

- LEXSY cell-free production of the to date largest multisubunit membrane protein complex<sup>(1)</sup> (A)
- Fluorescence-based protein interaction studies without purification<sup>(2,3,4)</sup>
- Fast high throughput screening<sup>(5,6,7)</sup>



Active HOPS membrane tethering complex (600 kDa) reconstituted *in vitro* from all six subunits co-expressed by *In vitro* LEXSY<sup>(1)</sup>.

- (1) Guo *et al.* (2013) *PLoS ONE* 8: e8153
- (2) Brooks *et al.* (2014) *Science* 344: 1249783
- (3) Gambin *et al.* (2014) *eLife* 3: e01434
- (4) Han *et al.* (2014) *Journal of Biological Chemistry* 289: 7764
- (5) Mureev *et al.* (2009) *Nature Biotechnology* 27: 747
- (6) Kovtun *et al.* (2010) *PLOS one* 5: e14388
- (7) Kovtun *et al.* (2011) *Methods* 55: 58

# LEXSY-expressed pharmaceutically relevant enzymes and diagnostics tools

## Pathogen diagnostics

- LEXSY-made surface antigens SAG1 and SAG2 of *Toxoplasma gondii* for ELISA kits<sup>(1)</sup>
- LEXSY-made PTX toxin for diagnostics of fungal-caused wheat disease<sup>(2)</sup>

## Antibodies

- Expression and purification of IgG based scFc fusions<sup>(3)</sup> from LEXSY
- LEXSY-expression and characterization of scFv collection<sup>(4)</sup>

## Pharma research

### LEXSY-made

- Proprotein convertase PC4 (subtilisin kexin) for inhibitor design<sup>(5)</sup>
- Human liver serine protease Coagulation Factor VII<sup>(6)</sup>
- Modified human Tissue-Plasminogen Activator (t-PA) that shows >100x higher activity than *E. coli* t-PA<sup>(7)</sup>
- Glycosylated Amyloid Precursor Protein sAPP $\alpha$  involved in Alzheimer disease<sup>(8)</sup>

(1) Ebert *et al.*, not published

(2) JBS, not published

(3) Jørgensen *et al.* (2014) *Microbial Cell Factories* 13: 9

(4) Klatt *et al.* (2012) *Microbial Cell Factories* 11: 97

(5) Basak *et al.* (2008) *Protein Expression and Purification* 60: 117

(6) Mirzaahmadi *et al.* (2011) *Journal of Biomedicine Biotechnology* 2011: 873874

(7) Nazari *et al.* (2011) *Biotechnology Letters* 33: 503

(8) Klatt *et al.* (2013) *Journal of Proteome Research* 12: 396



## Our LEXSY group...



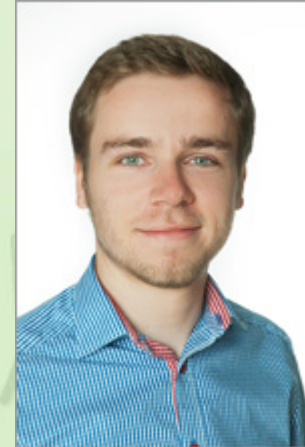
**Dr. Reinhard  
Breitling**  
„The LEXSYest Man  
Alive“



**Dr. Andreas  
Licht**



**Dr. Larissa  
Consani**  
Textor



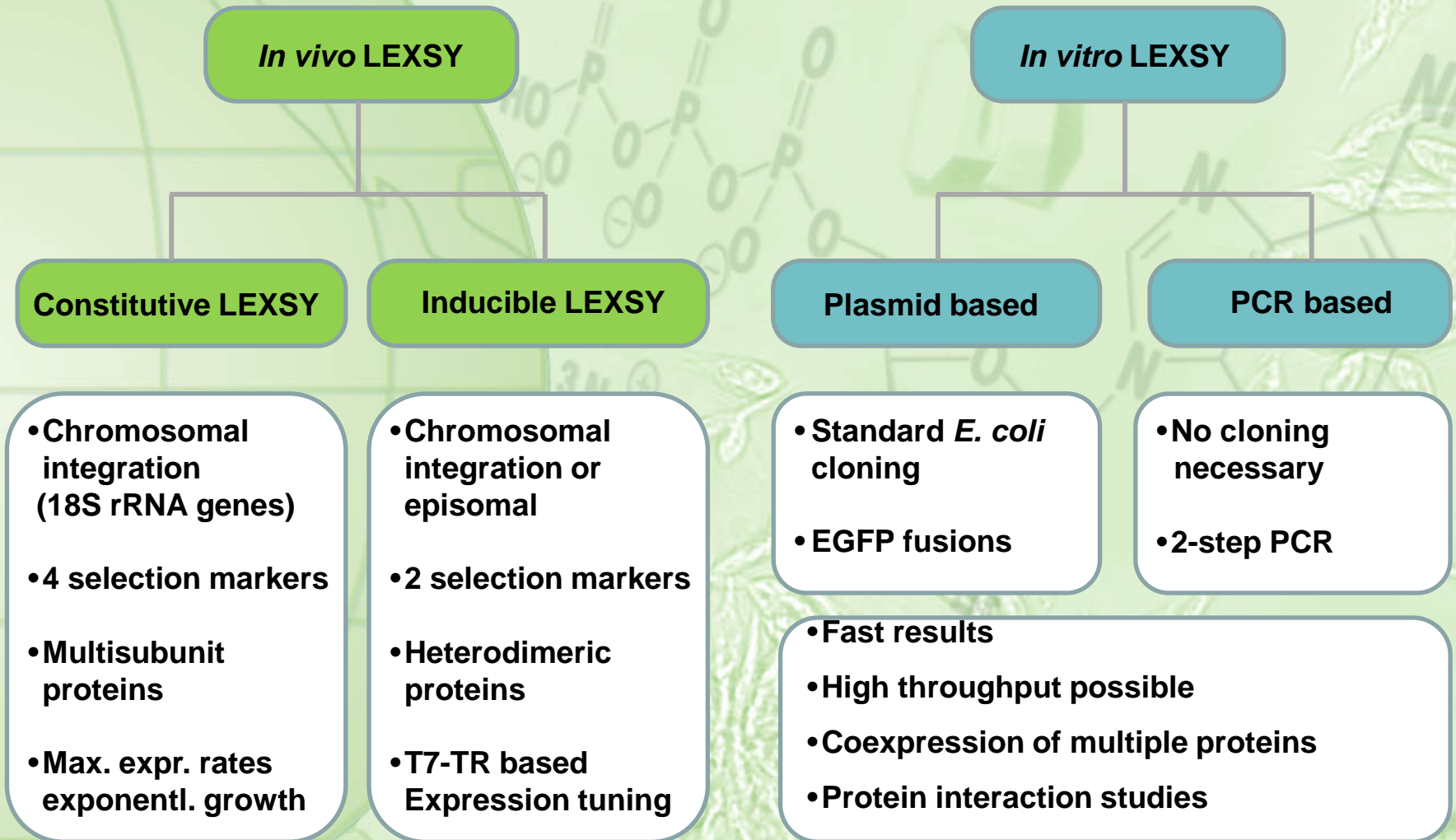
**Stefan  
Heiderich**

...answers any question when contacted at [expression@jenabioscience.com](mailto:expression@jenabioscience.com)

# backups



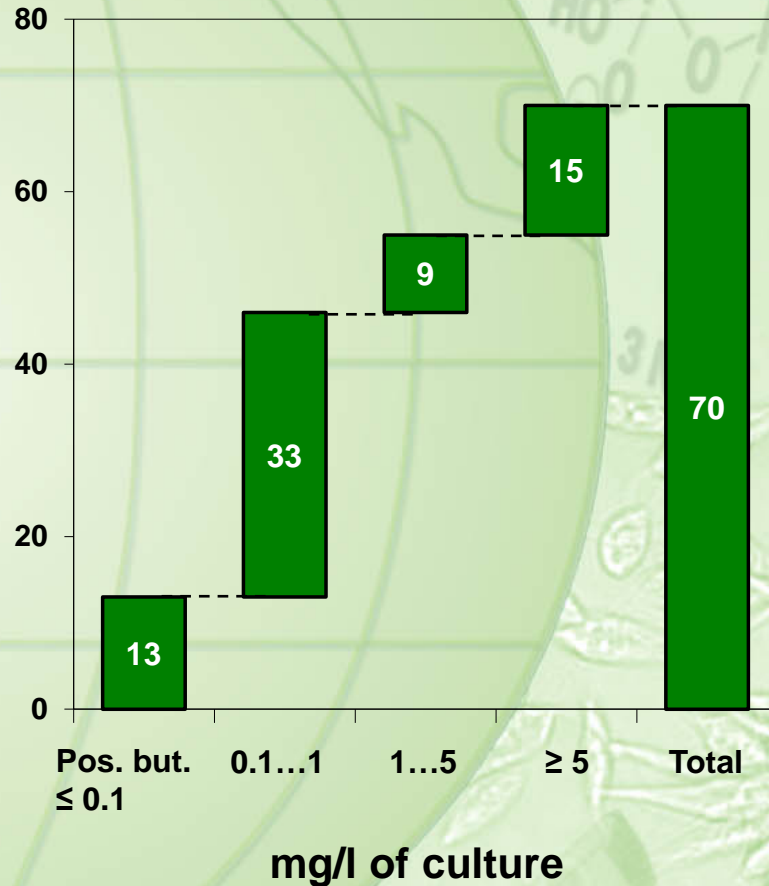
# Flexibility of LEXSY expression configurations provides solutions for your needs



# 80% successful (> 0.1 mg/l) expression projects

Random selection of 70 targets that gave unsatisfactory results in other expression systems

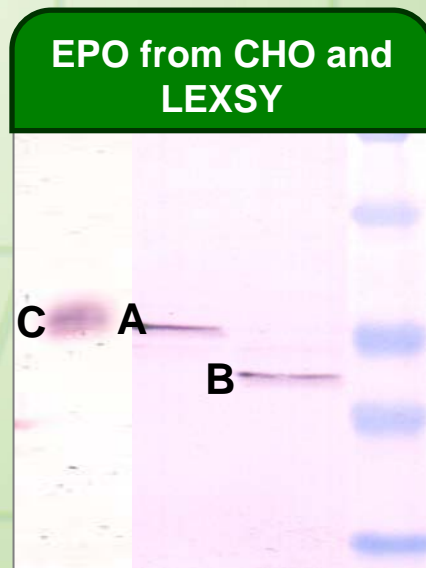
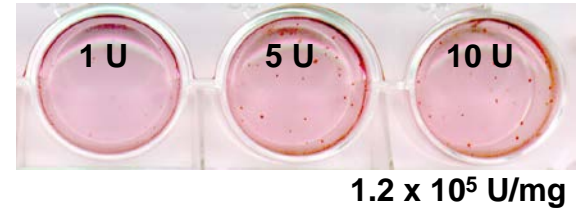
# of expression projects



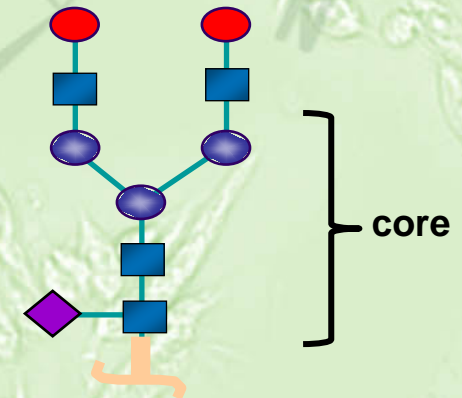
Target protein	Size kDa	Yield mg/L
<b>Cytoplasmic proteins</b>		
EGFP	28	300
SOD1	16	30
SPEE	35	30
p85 of PI3 kinase	85	3
smmyHC	154	1
<b>Nuclear proteins</b>		
T7 RNA Pol	100	1
<b>Secreted proteins</b>		
MHC II-β	30	500
CRP	23	44
SAG1&2	15/31	10
Fc fusion	39	10
MDP1	45	6
Laminin 332	420 (150+135+135)	0.5
<b>Membrane proteins</b>		
EGFP-Rab7 (mb-associated)	52	12
PDM9 (Type I)	43	0.5
BkrB2-GST (Type III TM7)	55	0.1

# Homogeneously glycosylated human Erythropoietin from LEXSY

- Completely secreted to the culture medium
- Natively processed at the N-terminus
- Biologically fully active
- Exceptionally homogeneously & mammalian-type N-glycosylated  
(biantennary fully galactosylated  $\text{Man}_3\text{GlcNAc}_2\text{core-}\alpha\text{-1,6-fucosylated}$  structure)



- A: homogeneously glycosylated EPO from LEXSY
- B: N-deglycosylated EPO from LEXSY
- C: heterogeneously glycosylated EPO from CHO



- Breitling *et al.* (2002) Non-pathogenic trypanosomatid protozoa as a platform for protein research and production. *Protein Expression and Purification* **25**: 209

# Full repertoire of eukaryotic PTMs in LEXSY

PTM	Homol. Targets <i>Leishmania</i>	Heterol. Targets LEXSY	Selected references
Disulfide bond formation (folding)	PMGP46	t-PA	Kahl <i>et al.</i> 1987, Hong <i>et al.</i> 2008 , Soleimani <i>et al.</i> 2007,
Multimerization	RNAP II, RNAP III	Laminin-322 SOD1, TETR	Phan <i>et al.</i> 2009, Gazdag <i>et al.</i> 2010, Kushnir <i>et al.</i> 2005, Martínez-Calvillo <i>et al.</i> 2007
Proteolytic processing - Signal peptide cleavage - Pro-region processing - Furin like processing	Gp63, 3'NT/NU CPB2.8	EPO, IFN $\gamma$ , SAG1 rPC4 sortilin	Breitling <i>et al.</i> 2002 and unpublished, Brooks <i>et al.</i> 2000, Basak <i>et al.</i> 2008 P. Madsen, not published
Glycosylation - N-Glycosylation - O-Glycosylation - Proteoglycan formation	gp63, sAP, 3'NT/NU sAP, KMP11 sAP	EPO, IFN $\gamma$ , SAG1	Breitling <i>et al.</i> 2002 and unpublished, Debrabant <i>et al.</i> 2000, Ellis <i>et al.</i> 2002, Ilg <i>et al.</i> 1994, Lippert <i>et al.</i> 1999, McGwire <i>et al.</i> 1996, Olafson <i>et al.</i> 1990, Stierhoff <i>et al.</i> 1998 Weise <i>et al.</i> 2000
Glypiation (GPI anchoring)	Gp63 Proteophosphoglycans MBAP <sup>GPI</sup>	rPC4 SAG1	McGwire <i>et al.</i> 1996, Ellis <i>et al.</i> 2002, Foth <i>et al.</i> 2002, Weise <i>et al.</i> (2000) , Basak <i>et al.</i> 2008, Breitling <i>et al.</i> unpublished,
Prenylation	LmLRAB, RAS-CVIM 7 non-assigned 14-140 kDa	Rab7	Chenik <i>et al.</i> 2006, Gillespie <i>et al.</i> 2007, Hasne <i>et al.</i> 1999, Alexandrov <i>et al.</i> not publ.

ARL-1 small G protein ADP-Ribosylation Factor-Like protein 1 of *L. donovani* , CPB2.8 Cysteine proteinase of *L. mexicana*, EPO hu Erythropoietin, Gp63 most abundant surface glycoprotein, GPI anchored and N-glycosylated HEXXH Zn metalloproteinase (leishmanolysin), H2A Histone 2A of *L. donovani*, HSP83-3 Heat shock protein of *L. donovani*, HSP60 Heat shock protein of *L. donovani*, IF3 Translation initiation factor 3 subunit of *L. donovani*, IFN $\gamma$  hu intereron gamma, KMP11 Kinetoplastid membrane protein of *L. donovani*, LIP2 60S acidic ribosomal protein P2 of *L. donovani*, LmLRAB RAB GTPase of *L. major*, Lt1200 1200 kDa Cytoskeletal Giant Protein of *L. tarentolae*, MBAP<sup>GPI</sup> artificially GPI anchored acid phosphatase of *L. mexicana*, 3'NT/NU Surface Membrane 3'-Nucleotidase/ Nuclease of *L. donovani*, OADC bacterial oxaloacetate decarboxylase in *L. mexicana amazonensis*, PMGP46 promastigote membrane glycoprotein of *L. mexicana amazonensis*, Rab7 geranylgeranyl transferase component A of *Rattus norvegicus*, Rbp16 RNA-binding protein of *L. donovani*, RNAP II RNA polymerase II of *L. major*, RNAP III RNA polymerase III of *L. major*, rPC4 rat proprotein convertase 4, S10 & S18 40S ribosomal proteins of *L. donovani*, SAG1 surface antigen 1 of *Toxoplasma gondii*, sAP secreted acid phosphatase (N- and O-glycosylated and phosphoglycosylation) of *L. mexicana* and *L. donovani*, SOD1 hu superoxide dismutase, TETR tetracycline repressor, t-PA hu tissue plasminogen activator, VG7A5 amastigote spec. Protein of *L. mexicana mexicana*

# Full repertoire of eukaryotic PTMs in LEXSY

PTM	Homol. Targets <i>Leishmania</i>	Heterol. Targets LEXSY	Selected references
Methylation	$\alpha$ -Tubulin (D329) Carboxypeptidase (E53) Rbp16 (R104), H2A (S97) ...		Rosenzweig <i>et al.</i> 2008, Jardim <i>et al.</i> 1995
Phosphorylation	Lt1200 (S,T), HSP83-3 (T) RNA helicase II (S) ...		Baqui <i>et al.</i> 2000, Rosenzweig <i>et al.</i> 2008
Acetylation - N-terminal  - internal	LIP2 (M1), KMP11 (A2), S10 (S2), IF3 (T2) ... S18 (S15), HSP60 (E293) ...	SOD1 (A2)	Rosenzweig <i>et al.</i> 2008, Gazdag <i>et al.</i> 2010, Thomas <i>et al.</i> 2009
Myristoylation	ARL-1		Sahin <i>et al.</i> 2008
Amidation	VG7A5		Liu <i>et al.</i> 2000
Glutathionylation	<i>Tb</i> mono-Cys-glutaredoxin 1		Melchers <i>et al.</i> 2007
ADP-ribosylation	TGN-Lysosome trafficking		Sturm <i>et al.</i> 1998, Sahin <i>et al.</i> 2008
Biotinylation (rec. strain)	-	OADC AVI-EGFP	Detke <i>et al.</i> 2007 Konthur <i>et al.</i> 2009

**ARL-1** small G protein ADP-Ribosylation Factor-Like protein 1 of *L. donovani*, **CPB2.8** Cysteine proteinase of *L. mexicana*, **EPO** hu Erythropoietin, **Gp63** most abundant surface glycoprotein, GPI anchored and N-glycosylated HEXxH Zn metalloproteinase (leishmanolysin), **H2A** Histone 2A of *L. donovani*, **HSP83-3** Heat shock protein of *L. donovani*, **HSP60** Heat shock protein of *L. donovani*, **IF3** Translation initiation factor 3 subunit of *L. donovani*, **IFN $\gamma$**  hu interferon gamma, **KMP11** Kinetoplastid membrane protein of *L. donovani*, **LIP2** 60S acidic ribosomal protein P2 of *L. donovani*, **LmLRAB** RAB GTPase of *L. major*, **Lt1200** 1200 kDa Cytoskeletal Giant Protein of *L. tarentolae*, **MBAP<sup>GPI</sup>** artificially GPI anchored acid phosphatase of *L. mexicana*, **3'NT/NU** Surface Membrane 3'-Nucleotidase/ Nuclease of *L. donovani*, **OADC** bacterial oxaloacetate decarboxylase in *L. mexicana amazonensis*, **PMGP46** promastigote membrane glycoprotein of *L. mexicana amazonensis*, **Rab7** geranylgeranyl transferase component A of *Rattus norvegicus*, **Rbp16** RNA-binding protein of *L. donovani*, **RNAP II** RNA polymerase II of *L. major*, **RNAP III** RNA polymerase III of *L. major*, **rPC4** rat proprotein convertase 4, **S10 & S18** 40S ribosomal proteins of *L. donovani*, **SAG1** surface antigen 1 of *Toxoplasma gondii*, **sAP** secreted acid phosphatase (N- and O-glycosylated and phosphoglycosylation) of *L. mexicana* and *L. donovani*, **SOD1** hu superoxide dismutase, **TETR** tetracycline repressor, **t-PA** hu tissue plasminogen activator, **VG7A5** amastigote spec. Protein of *L. mexicana mexicana*

The background is a light green collage. On the left, a semi-transparent globe shows latitude and longitude lines. Scattered throughout are various scientific illustrations: a complex phosphate chain with three phosphorus atoms (P) and several oxygen atoms (O), some with negative charges; a nucleotide structure with a nitrogenous base (containing N and NH2 groups) attached to a sugar ring (with OH groups); and several biological cells, including what appear to be plant cells with cell walls and chloroplasts, and smaller, more irregular cells.

**unused**



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